

## Review

# Ginger—An Herbal Medicinal Product with Broad Anti-Inflammatory Actions

Reinhard Grzanna,<sup>1</sup> Lars Lindmark,<sup>2</sup> and Carmelita G. Frondoza<sup>3</sup>

<sup>1</sup>RMG Biosciences, Inc.; <sup>3</sup>Department of Orthopaedic Surgery, Johns Hopkins University School of Medicine, Baltimore, Maryland; and <sup>2</sup>Ferrosan A/S, Soeborg, Denmark

**ABSTRACT** The anti-inflammatory properties of ginger have been known and valued for centuries. During the past 25 years, many laboratories have provided scientific support for the long-held belief that ginger contains constituents with anti-inflammatory properties. The original discovery of ginger's inhibitory effects on prostaglandin biosynthesis in the early 1970s has been repeatedly confirmed. This discovery identified ginger as a herbal medicinal product that shares pharmacological properties with non-steroidal anti-inflammatory drugs. Ginger suppresses prostaglandin synthesis through inhibition of cyclooxygenase-1 and cyclooxygenase-2. An important extension of this early work was the observation that ginger also suppresses leukotriene biosynthesis by inhibiting 5-lipoxygenase. This pharmacological property distinguishes ginger from non-steroidal anti-inflammatory drugs. This discovery preceded the observation that dual inhibitors of cyclooxygenase and 5-lipoxygenase may have a better therapeutic profile and have fewer side effects than non-steroidal anti-inflammatory drugs. The characterization of the pharmacological properties of ginger entered a new phase with the discovery that a ginger extract (EV.EXT.77) derived from *Zingiber officinale* (family Zingiberaceae) and *Alpinia galanga* (family Zingiberaceae) inhibits the induction of several genes involved in the inflammatory response. These include genes encoding cytokines, chemokines, and the inducible enzyme cyclooxygenase-2. This discovery provided the first evidence that ginger modulates biochemical pathways activated in chronic inflammation. Identification of the molecular targets of individual ginger constituents provides an opportunity to optimize and standardize ginger products with respect to their effects on specific biomarkers of inflammation. Such preparations will be useful for studies in experimental animals and humans.

**KEY WORDS:** • chemokines • cyclooxygenase • cytokines • ginger • inflammation

## INTRODUCTION

**G**INGER (*Zingiber officinale* Roscoe) has a long history of medicinal use.<sup>1,2</sup> In traditional Chinese and Indian medicine, ginger has been used to treat a wide range of ailments including stomachaches, diarrhea, nausea, asthma, respiratory disorders, toothache, gingivitis, and arthritis.<sup>2,3</sup> Today, ginger and its extracts are recommended by herbal practitioners primarily for dyspepsia and the prevention of motion sickness.<sup>4</sup> A number of recent studies have renewed interest in ginger for the treatment of chronic inflammatory conditions. This interest can be traced to the discovery in the early 1970s that non-steroidal anti-inflammatory drugs (NSAIDs) produce their effects by inhibiting the biosyn-

thesis of prostaglandins (PGs).<sup>5</sup> Soon thereafter, ginger was found to contain constituents that inhibit PG synthesis, a finding that provided a sound scientific rationale for its anti-inflammatory effects.<sup>6</sup> Subsequent studies revealed that ginger also contains constituents with pharmacological properties similar to the novel class of dual-acting NSAIDs.<sup>7</sup> Compounds in this class inhibit arachidonic acid metabolism via the cyclooxygenase (COX) and lipoxygenase (LOX) pathways. These compounds have notably fewer side effects than conventional NSAIDs and now are being investigated as a novel class of anti-inflammatory compounds.<sup>8–10</sup>

Results of recent studies summarized in this review have shown that ginger's pharmacological effects on the inflammatory process extend well beyond the inhibition of PG synthesis. These studies uncovered an effect of ginger on the production of cytokines that are synthesized and secreted at sites of inflammation. These molecules have become highly promising targets for the treatment of chronic inflammatory disorders.<sup>11</sup> The preclinical findings on mechanisms by

Manuscript received 12 August 2004. Revision accepted 11 November 2004.

Address reprint requests to: Carmelita G. Frondoza, Ph.D., Department of Orthopaedic Surgery, Johns Hopkins University School of Medicine, Good Samaritan Hospital, 5601 Loch Raven Boulevard, Baltimore, MD 21239, E-mail: cgfrondo@jhmi.edu

which ginger produces its effects are complemented by recent clinical trials that support the traditional view that ginger has analgesic and anti-inflammatory properties.<sup>12–15</sup> Results from clinical trials, observational studies, and case reports on the medicinal use of ginger can be found on the Internet.<sup>12</sup> This review contains an account of the scientific data on ginger compiled over the past 25 years. These data strongly support the commonly held view that ginger is a valuable medicinal product for the treatment of chronic inflammatory conditions.

## CONSTITUENTS OF GINGER

As is the case with many other herbal preparations, ginger extracts are a complex, multicomponent mixture of biologically active constituents. More than 400 chemical compounds have been isolated and identified in extracts of ginger rhizomes, and new ones are still being detected.<sup>16–19</sup> At present, only a few of them have been evaluated for their pharmacological properties. Current evidence suggests that a subfraction containing the structurally related compounds gingerols, shogaols, and paradols accounts for a major portion of ginger's anti-inflammatory properties. These compounds share an aromatic ketone moiety but differ in the length of their alkyl side chain and the substitution pattern on the side chain. Structure–activity relationship analysis suggests that the presence of the phenolic hydroxy group adjacent to the methoxy group is critical for the inhibition of PG synthesis.<sup>20,21</sup> The relative proportions of gingerols, shogaols, and paradols in ginger extracts are determined by a number of factors, including the geographic origin, the maturity of the rhizomes at the time of harvest, and the method by which the extracts are prepared. Shogaols, dehydrated products of gingerols, are a major component of dried ginger powder. Gingerols are thermally labile because of the presence of a  $\beta$ -hydroxy keto group, which readily undergoes dehydration to form the corresponding shogaols (Fig. 1). Shogaols may be further converted to paradols by hydrogenation. Shogaols and zingerone are found only in small quantities in fresh ginger, but are present in large amounts in stored ginger, suggesting that this conversion takes place during processing and storage. The extent of this conversion is likely to have a significant impact on the health effects of ginger preparations since the two classes of compounds are likely to vary in their bioavailability, pharmacokinetics, and pharmacological properties.

Differences in the methods by which ginger extracts are prepared make it difficult to compare the results of studies from various laboratories. Comparative studies of the pharmacological properties of ginger would be greatly facilitated if extracts would be standardized in reference to a universally accepted, internal constituent. Since the inhibitory effects of ginger on PG synthesis can be attributed to the presence of hydroxymethoxyphenyl compounds (HAPC), it is reasonable to use these compounds as an internal standard. The ginger preparation EV.EXT.77, which is derived from *Z. officinale* Roscoe and *Alpina galanga*, is standardized in reference to HAPC and acetoxychavicol acetate (ACA)

compounds. HAPC include all constituents containing the 3-methoxy-4-hydroxyphenyl moiety (mainly gingerols and shogaols). According to analytical certificates, each capsule of EV.EXT.77 contains a minimum of 30 mg of HAPC/ACA in 255 mg of mixed extract complex. Despite the success in tracking the anti-inflammatory activities in chromatographic fractions of ginger extracts,<sup>6,21</sup> no attempts have been made to standardize preparations based on bioactivity. This method would have the advantage that it does not require knowledge of the identity of pharmacologically active constituents.<sup>22</sup> This approach would permit mixtures derived from different sources to be standardized with respect to an accepted biological readout such as inhibition of PG synthesis. Such biological assay would be especially useful if active constituents of ginger exert their anti-inflammatory effects via different pharmacological mechanisms and thus may act synergistically.

## GINGER INHIBITS PG BIOSYNTHESIS THROUGH INHIBITION OF COX-1 AND COX-2 ENZYME ACTIVITY

A major breakthrough in inflammation research was the discovery by Vane<sup>5</sup> in 1971 that aspirin and related drugs produce their anti-inflammatory effects by inhibiting the synthesis of PGs. This seminal observation prompted several laboratories to explore whether naturally occurring substances with known anti-inflammatory properties also act as inhibitors of PG synthesis. Kiuchi *et al.*<sup>6</sup> were the first to show that extracts of plants belonging to the Zingiberaceae family inhibit PG synthesis *in vitro*. These investigators subjected extracts of fresh ginger to chromatographic purification and analyzed the resulting fractions for their effect on PG synthesis. They isolated and identified [6]-gingerol and

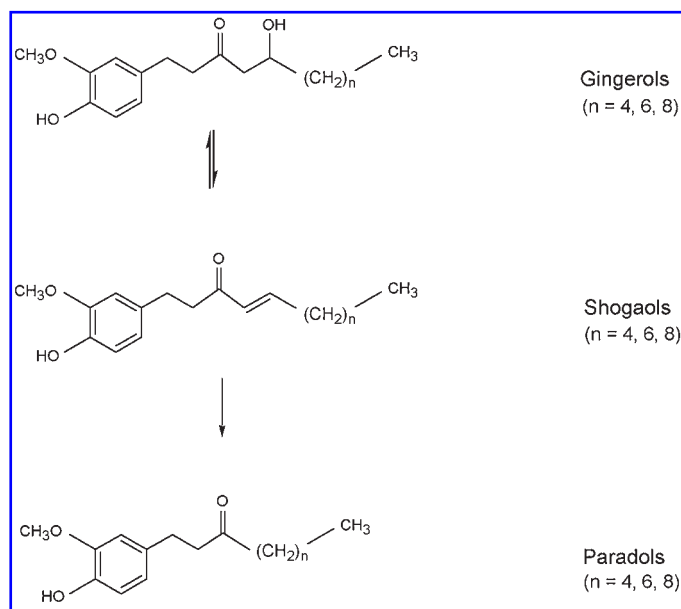


FIG. 1. Chemical structures of some ginger constituents.

four structurally related compounds that inhibit PG synthesis in rabbit renal medulla homogenates with  $IC_{50}$  values ranging from 1.0 to 5.5  $\mu M$ . Under the same assay conditions, indomethacin, one of the most potent inhibitors of PG synthesis, had an  $IC_{50}$  value of 4.9  $\mu M$ . These studies provided the first direct experimental evidence that ginger contains several constituents with anti-inflammatory activities comparable in potency to NSAIDs.

The inhibition of PG synthesis by NSAIDs and ginger is due to inhibition of arachidonic acid metabolism by COX.<sup>23</sup> This enzyme exists in at least two distinct isoforms, designated COX-1 and COX-2.<sup>24</sup> COX-1 is constitutively expressed in nearly all cells and tissues. It regulates important physiologic processes such as gastrointestinal cytoprotection and electrolyte homeostasis in kidneys. In contrast, COX-2 is almost undetectable in most tissues, but its expression is greatly increased at sites of inflammation.<sup>24</sup> It is generally accepted that many of the toxic effects of NSAIDs are due to inhibition of COX-1, while the therapeutic effects reside in the inhibition of COX-2. These observations have led to major efforts by pharmaceutical companies to develop NSAIDs that preferentially inhibit COX-2.<sup>25</sup> So far, only one study has attempted to determine the relative potency of gingerols on COX enzyme activity in intact cells. A study by Tjendraputra *et al.*<sup>20</sup> showed that gingerols are somewhat more potent inhibitors of COX-1 than COX-2, thus identifying them as non-selective COX inhibitors. This was a surprising finding since non-selective NSAIDs are known for their gastrointestinal and renal side effects.<sup>26</sup> In fact, ginger extracts have anti-ulcer activity and are recommended for the treatment of gastrointestinal problems.<sup>26–28</sup> The lack of ginger's gastrointestinal side effects suggested the presence of a yet unidentified pharmacological activity responsible for the protective effects against the toxicity associated with COX-1 inhibition.

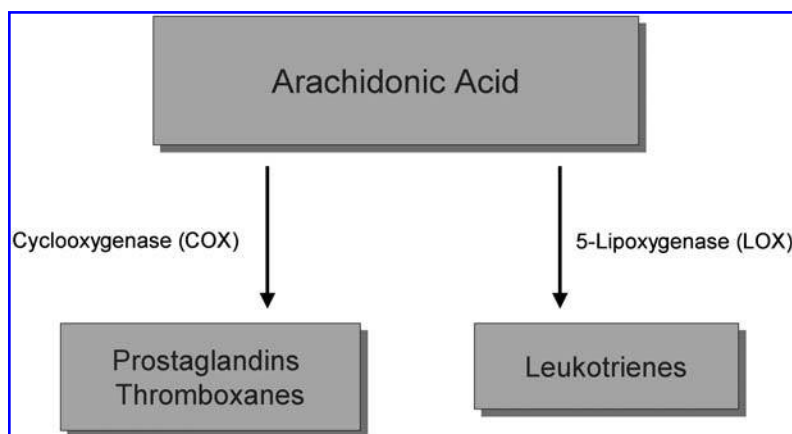
### GINGER CONSTITUENTS ARE DUAL INHIBITORS OF COX AND 5-LOX

An important advance in the characterization of the anti-inflammatory properties of ginger was the discovery that some of its constituents not only inhibit PG synthesis but

also leukotriene (LT) synthesis. LTs are derived from arachidonic acid through the action of the enzyme 5-LOX (Fig. 2). LTs are potent mediators of the inflammatory process and are suspected of playing a key role in the development of gastrointestinal ulcers associated with long-term use of NSAIDs.<sup>29,30</sup> Flynn and Rafferty<sup>7</sup> were the first to show that several ginger constituents are dual inhibitors of arachidonic acid metabolism. Compounds containing the 4-hydroxy-3-methoxyphenyl moiety, including [6]-gingerol, shagaol, gingerdione, and dihydroparadol, inhibit PG and LT production in human neutrophils in the low micromolar range. This observation prompted Flynn and Rafferty<sup>7</sup> to designate some ginger constituents as functional dual inhibitors of COX and LOX. Kiuchi *et al.*<sup>21</sup> found gingerols with long alkyl side chains ( $n > 6$ ; see Fig. 1) to be more potent inhibitors of LT synthesis than of PG synthesis. The significance of this observation was not fully appreciated until the recent discovery that dual inhibitors of COX and LOX are more effective and have fewer gastrointestinal side effects than pure COX inhibitors.<sup>8,10</sup> Nickerson-Nutter and Medvedoff<sup>31</sup> showed that inhibitors of PG and LT synthesis in combination are more effective in preventing collagen-induced arthritis in animals than inhibitors of either class of compounds alone. Unlike ginger, NSAIDs do not inhibit the synthesis of LTs from arachidonic acid. Consequently, COX inhibition by NSAIDs shifts arachidonic acid metabolism to the LOX pathway, leading to an increased production of LTs.<sup>32</sup> Compounds with this dual effect on COX and LOX are being evaluated as a new class of anti-inflammatory drugs.<sup>8–10</sup> The observation that constituents of ginger are dual COX/LOX inhibitors may explain why even high doses of ginger extract do not produce the side effects often observed with non-selective COX inhibitors.

### GINGER HAS A HISTORY OF USE FOR THE TREATMENT OF RHEUMATISM

Several clinical studies support the value of ginger for the treatment of arthritis. In addition to alleviating pain, ginger extract has been reported to decrease joint swelling.<sup>33,34</sup> In an exploratory trial of 57 osteoarthritic cases the ginger extract EV.EXT.33 prepared from the rhizomes of *Z. officinalis*



**FIG. 2.** Arachidonic acid is metabolized via two separate pathways: Metabolism along the COX pathway leads to the formation of PGs and thromboxanes, while metabolism along the LOX pathway leads to the formation of LTs.

*nale* was reported to be better than placebo during the first period of treatment before cross-over with NSAIDs.<sup>14</sup> Altman and Marcussen<sup>13</sup> using a similar ginger extract (EV.EXT.77) conducted a 6-week, double-blind placebo-controlled parallel group study involving 247 patients with osteoarthritis and demonstrated a statistically significant reduction in knee pain. Their analysis of secondary efficacy variables also showed a consistently superior response in patients treated with ginger extract compared with the control group. Another 6-month, double-blind placebo-controlled study of 29 osteoarthritic patients evaluated the ginger extract Zintona EC derived from *Z. officinale*; this study also observed significantly reduced knee pain compared with controls.<sup>15</sup> These recent clinical studies confirm the beneficial effects of ginger extracts in the treatment of arthritis and inflammation.

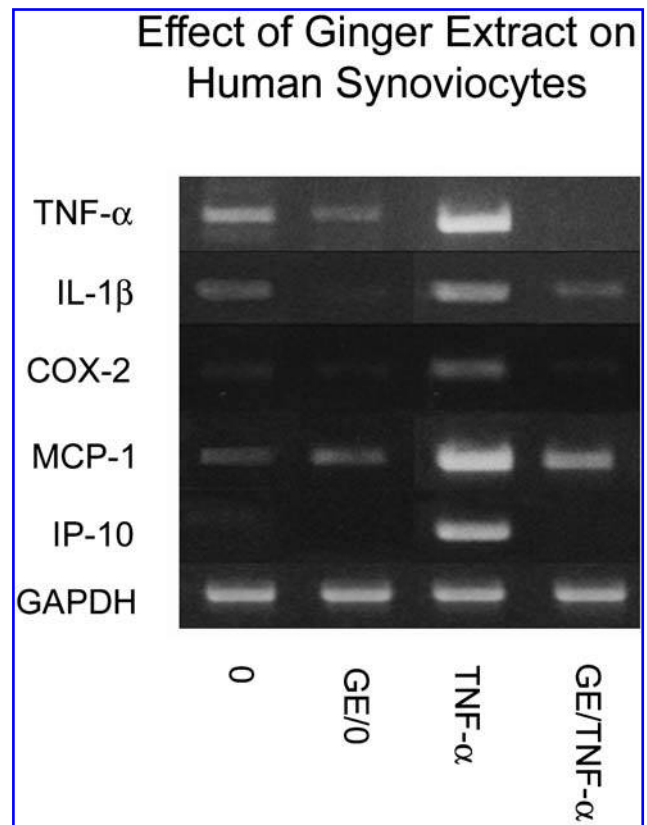
### GINGER EXTRACT INHIBITS CYTOKINE AND CHEMOKINE INDUCTION IN SYNOVIOCYTES AND CHONDROCYTES—IMPLICATIONS FOR THE TREATMENT OF ARTHRITIS

Genes encoding pro-inflammatory cytokines are up-regulated in chronic inflammatory conditions. Cytokines are a class of small proteins that are secreted at sites of inflammation principally by lymphocytes, macrophages, and fibroblasts. They function as chemical messengers between cells involved in immune and inflammatory responses. Chemokines are a subset of cytokines that act primarily as chemoattractants by inducing the recruitment of effector cells to sites of tissue damage. Inhibiting the production of pro-inflammatory cytokines or blocking their actions is a new and successful therapeutic approach for the treatment of inflammatory disorders,<sup>11</sup> most notably rheumatoid arthritis.<sup>35</sup> Several studies have shown that a number of natural products also contain constituents capable of inhibiting cytokine induction in cells.<sup>36</sup> For example, the flavonoid wogonin inhibits inflammatory activation of microglial cells in culture by diminishing lipopolysaccharide (LPS)-induced tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin (IL)-1 $\beta$ , and nitric oxide induction.<sup>37</sup> Chang *et al.*<sup>38</sup> demonstrated that wogonin inhibits monocyte chemotactic protein-1 gene expression in human endothelial cells. Some natural products of the terpenoid family have been reported to suppress inflammatory processes via inhibition of the production of TNF- $\alpha$  and IL-1 $\beta$ .<sup>39–41</sup> Several products are available or are currently under development for controlling the adverse effects of TNF- $\alpha$  and IL-1 $\beta$  overproduction in rheumatoid arthritis. These include diacerein, a drug with IL-1 inhibitory activity *in vitro*,<sup>42</sup> etanercept, a TNF- $\alpha$  receptor fusion protein that competitively inhibits the binding of TNF- $\alpha$  to its receptor, and infliximab, a monoclonal antibody that blocks human TNF- $\alpha$  receptors.<sup>11,35</sup>

Osteoarthritis is associated with a variable degree of inflammation. Activation of synovial cells in joints leads to the release of TNF- $\alpha$  and IL-1 $\beta$ .<sup>43</sup> To determine whether a

ginger extract can inhibit the induction of pro-inflammatory cytokines, we have conducted a series of experiments in human synoviocytes and chondrocytes.<sup>44,45</sup> Synoviocytes obtained during arthroplasty from osteoarthritic patients were activated with either TNF- $\alpha$  or IL-1 $\beta$ , the two key cytokines involved in the inflammation and degradation of joints. Ginger extract inhibited the expression of TNF- $\alpha$  in synoviocytes activated by either IL-1 $\beta$  or TNF- $\alpha$  at the transcript (Fig. 3) and protein levels.<sup>46</sup> Similarly, ginger extract suppressed TNF- $\alpha$  expression in activated chondrocytes.<sup>45</sup> The ginger extract also inhibited the induction of genes encoding chemokines: monocyte chemotactic protein-1 and interferon- $\gamma$  inducible protein-10 (see Fig. 3).<sup>47</sup>

In addition to induction of cytokines and chemokines, COX-2 protein levels are also greatly increased in inflamed joint tissue because of COX-2 gene induction.<sup>48</sup> Overexpression of COX-2 is a characteristic feature of osteoarthritis,<sup>49</sup> rheumatoid arthritis,<sup>50</sup> and a number of other pathological conditions such as atherosclerosis, inflammatory



**FIG. 3.** Inhibition of cytokine and chemokine induction by ginger extract (GE) in human synoviocytes. Synoviocytes were incubated for 1 hour with media in the absence (lane 1) or presence of 100  $\mu$ g/mL of GE (lane 2). Non-activated human synoviocytes express moderate levels of TNF- $\alpha$  and low levels of IL-1 $\beta$ . When treated with TNF- $\alpha$  (1 ng/mL) for 1 hour, synoviocytes in media alone showed increased levels of TNF- $\alpha$  and IL-1 $\beta$  mRNA (lane 3). These increases were not observed in cultures of synoviocytes pretreated with GE (lane 4). GAPDH, glyceraldehyde phosphate dehydrogenase; IP-10, interferon- $\gamma$  inducible protein-10; MCP-1, monocyte chemotactic protein-1.





## GINGER INHIBITS THE TRANSCRIPTION FACTOR NUCLEAR FACTOR- $\kappa$ B (NF- $\kappa$ B)

NF- $\kappa$ B is a principal regulator of pro-inflammatory gene expression.<sup>68</sup> These include genes encoding cytokines, chemokines, and the enzyme COX-2.<sup>69</sup> Activated NF- $\kappa$ B can be detected at sites of inflammation, and a link among NF- $\kappa$ B activation, cytokine production, and inflammation is now generally accepted.<sup>70</sup> Aberrant NF- $\kappa$ B activation has been observed in synovial tissues of both osteoarthritis and rheumatoid arthritis<sup>71</sup> and in several other chronic inflammatory diseases.<sup>72</sup> NF- $\kappa$ B-directed therapies have been shown to be effective in several animal model of inflammatory diseases.<sup>70</sup> A number of natural products known for their anti-inflammatory properties are now suspected to produce their effects through inhibition of the NF- $\kappa$ B pathway.<sup>36</sup> Several naturally occurring phenolic compounds including curcumin and flavonoids have already been shown to inhibit NF- $\kappa$ B. Based on the observation that ginger extract inhibits pro-inflammatory gene expression, Frondoza *et al.*<sup>46</sup> tested the effect of the ginger extract EV.EXT.77 on NF- $\kappa$ B expression *in vitro*. The results show that this ginger extract significantly inhibits NF- $\kappa$ B expression in activated synoviocytes at 100  $\mu$ g/mL. Such a mechanism would offer an explanation for the broad effect of ginger on inflammatory processes in various cell types and tissues.

## GINGEROLS AS VANILLOID RECEPTOR AGONISTS—A NOVEL MECHANISM FOR GINGER'S EFFECT ON PAIN

The chemical mediators released by local or recruited cells stimulate nociceptive primary afferents at sites of inflammation resulting in pain.<sup>73,74</sup> Joint inflammation in chronic arthritis leads to hyperalgesia, a state characterized by an excessive response to noxious stimuli. PGs, TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and IL-8 all contribute to acute and chronic pain associated with inflammation. NSAIDs are effective in the management of inflammation-related pain. Similarly, the effects of ginger in reducing joint pain are also likely to be mediated through its effect on PG synthesis and other inflammatory mediators. Recently, a novel molecular target of ginger constituents was identified, suggesting an additional mechanism by which ginger may reduce inflammatory pain. Dedov *et al.*<sup>75</sup> reported that gingerols act as agonists at vanilloid receptors. These receptors had previously been identified as receptors activated by capsaicin and are suspected to be present on pain afferents mediating joint pain.<sup>76</sup> Vanilloid receptor agonists are potent analgesics, and the finding by Dedov *et al.*<sup>75</sup> has added gingerols and zingerone to the list of vanilloid receptor agonists. The discovery that ginger constituents function as agonists at vanilloid receptors provides a new mechanistic explanation for the well-documented analgesic effect of ginger in the treatment of pain in rheumatic and inflammatory conditions.

## CONCLUSION

A considerable body of scientific data supports the long-held view that ginger has a broad spectrum of anti-inflammatory activities. The scientific data reviewed here show that ginger produces its anti-inflammatory effects through multiple mechanisms. Ginger shares with NSAIDs the property of inhibiting PG synthesis. Some ginger constituents are dual inhibitors of COX and LOX, and thereby reduce the biosynthesis of both PGs and LTs. This remarkable property distinguishes ginger from conventional NSAIDs and may account for its lack of gastrointestinal and renal side effects. The recently discovered anti-cytokine effects of ginger show that the anti-inflammatory properties of this commonly used herb are far from being fully characterized. The identity of the constituents accounting for ginger's anti-cytokine effects remains to be determined. Future studies will need to address the question whether the anti-inflammatory effects of ginger so well documented in *in vitro* experiments can be verified in animals and humans. The results of clinical studies conducted so far have been encouraging. Considering the broad spectrum of ginger's anti-inflammatory actions and its safety record, this herbal product is likely to be a valuable dietary supplement in the treatment of inflammatory disorders.

## ACKNOWLEDGMENTS

Some of the experiments reviewed here were supported by a grant from the National Cancer Institute, National Institutes of Health and by Ferrosan A/S.

## REFERENCES

1. Afzal M, Al-Hadidi D, Menon M, Pesek M, Dhami MSI: Ginger: An ethnomedical, chemical and pharmacological review. *Drug Metab Drug Interact* 2001;18:159–190.
2. Awang DVC: Ginger. *Can Pharmaceut J* 1992;125:309–311.
3. Leung AY: *Encyclopedia of Common Natural Ingredients Used in Food, Drugs and Cosmetics*, John Wiley and Sons, New York, 1980.
4. Blumenthal M: *The Complete German Commission E Monographs*, American Botanical Council, Boston, 1998.
5. Vane JR: Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nature New Biol* 1971;231:232–235.
6. Kiuchi F, Shibuya M, Sankawa U: Inhibitors of prostaglandin biosynthesis from ginger. *Chem Pharm Bull (Tokyo)* 1982;30:754–757.
7. Flynn DL, Rafferty MF: Inhibition of human neutrophil 5-lipoxygenase activity by gingerdione, shagaol, capsaicin and related pungent compounds. *Prostaglandins Leukot Med* 1986;24:195–198.
8. Celotti F, Laufer S: Anti-inflammatory drugs: New multitarget compounds to face an old problem. The dual inhibition concept. *Pharmacol Res* 2001;43:429–436.
9. Fiorucci S, Meli R, Bucci M, Cirino G: Dual inhibitors of cyclooxygenase and 5-lipoxygenase. A new avenue to anti-inflammatory therapy? *Biochem Pharmacol* 2001;62:1433–1438.

10. Martel-Pelletier J, Lajeunesse D, Reboul P, Pelletier J-P: Therapeutic role of dual inhibitors of 5-LOX and COX, selective and non-selective non-steroidal anti-inflammatory drugs. *Ann Rheum Dis* 2003;62:501–509.
11. Dinarello CA: Anti-cytokine therapeutics and infections. *Vaccine* 2003;21(Suppl):S2/24–S22/34.
12. <http://www.herbmed.org/herbs/herb138.htm> (accessed 18 April 2005).
13. Altman RD, Marcussen KC: Effects of a ginger extract on knee pain in patients with osteoarthritis. *Arthritis Rheum* 2001;44:2461–2462.
14. Bliddal H, Rosetzky A, Schlichting P, Weidner MS, Andersen LA, Ibfelt H-H, Christensen K, Jensen ON, Barsley J: A randomized, placebo-controlled, cross-over study of ginger extracts and ibuprofen in osteoarthritis. *Osteoarthritis Cartilage* 2000;8:9–12.
15. Wigler I, Grotto I, Caspi D, Yaron M: The effects of Zintona EC (a ginger extract) on symptomatic gonarthrosis. *Osteoarthritis Cartilage* 2003;11:783–789.
16. Duke JA: *Handbook of Medicinal Herbs*, CRC Press, Boca Raton, FL, 2002.
17. Ma J, Jin XJ, Yang L, Liu Z-L: Diarylheptanoids from the rhizomes of *Zingiber officinale*. *Phytochemistry* 2004;65:1137–1143.
18. Jolad SD, Lantz RC, Solyom AM, Chen GJ, Bates RB, Timmermann BN: Fresh organically grown ginger (*Zingiber officinale*): Composition and effects on LPS-induced PGE<sub>2</sub> production. *Phytochemistry* 2004;65:1937–1954.
19. Charles R, Garg SN, Kumar S: New gingerdione from the rhizomes of *Zingiber officinale*. *Fitoterapia* 2000;71:716–718.
20. Tjendraputra E, Tran VH, Liu-Brennan D, Roufogalis BD, Duke CC: Effect of ginger constituents and synthetic analogues on cyclooxygenase-2 enzyme in intact cells. *Bioorg Chem* 2001;29:156–163.
21. Kiuchi F, Iwakami S, Shibuya M, Hanaoka F, Sankawa U: Inhibition of prostaglandin and leukotriene biosynthesis by gingerols and diaryl heptanoids. *Chem Pharm Bull (Tokyo)* 1992;40:387–391.
22. McLaughlin JL, Rogers LL, Anderson JE: The use of biological assays to evaluate botanicals. *Drug Inform J* 1998;32:513–524.
23. Srivastava K: Isolation and effects of some ginger compounds on platelet aggregation and eicosanoid biosynthesis. *Prostaglandins Leukot Med* 1986;25:187–198.
24. Vane JR, Bakhele YS, Botting RM: Cyclooxygenases I and 2. *Annu Rev Pharmacol Toxicol* 1998;38:97–120.
25. Warner TD, Mitchell JA: Cyclooxygenases: New forms, new inhibitors, and lessons from the clinic. *FASEB J* 2004;18:790–804.
26. Sertie JAA, Basile AC, Oshiro TT, Silva FD, Mazella AAG: Preventive anti-ulcer activity of the rhizome extract of *Zingiber officinale*. *Fitoterapia* 1992;63:55–59.
27. Yoshikawa M, Yamagashi S, Kumini K, Matsuda H, Okuno Y, Yamashara J, Murakami A: Stomachic principles in ginger. An anti-ulcer principle 6-gingesulfonic acid, and three monoacyldigalactosylglycerols gingerglycolipids A, B, and C, from *Zingiber* rhizome originating in Taiwan. *Chem Pharm Bull (Tokyo)* 1994;42:226–230.
28. Yamahara J, Mochizuki M, Rong HQ, Matsuda H, Fujimura H: The anti-ulcer effect in rats of ginger constituents. *J Ethnopharmacol* 1988;23:299–304.
29. Hudson N, Balsitis M, Everitt S, Hawkey CJ: Enhanced gastric mucosal leukotriene B<sub>4</sub> synthesis in patients taking non-steroidal antiinflammatory drugs. *Gut* 1993;34:742–747.
30. Asako H, Kubes P, Wallace J, Wolf RE, Granger DN: Indomethacin-induced leukocyte adhesion in mesenteric venules: Role of lipoxygenase products. *Am J Physiol* 1992;262:G903–G908.
31. Nickerson-Nutter CL, Medvedeff ED: The effect of leukotriene synthesis inhibitors in models of acute and chronic inflammation. *Arthritis Rheum* 1996;39:515–521.
32. Laufer S: Role of eicosanoids in structural degradation in osteoarthritis. *Curr Opin Rheumatol* 2003;15:623–627.
33. Srivastava KC, Mustafa T: Ginger (*Zingiber officinale*) and rheumatic disorders. *Med Hypotheses* 1989;29:25–28.
34. Srivastava K, Mustafa T: Ginger (*Zingiber officinale*) in rheumatism and musculoskeletal disorders. *Med Hypotheses* 1992;39:342–348.
35. Choy EHS, Panayi GS: Cytokine pathways and joint inflammation in rheumatoid arthritis. *N Engl J Med* 2001;344:907–916.
36. Bremner P, Heinrich M: Natural products as targeted modulators of the nuclear factor- $\kappa$ B pathway. *J Pharm Pharmacol* 2001;54:453–472.
37. Lee H, Kim YO, Kim H, Kim SY, Noh HS, Kang SS, Cho GJ, Choi WS, Suk K: Flavonoid wogonin from medicinal herb is neuroprotective by inhibiting inflammatory activation of microglia. *FASEB J* 2003;17:1943–1944.
38. Chang Y-L, Shen J-J, Wung B-S, Cheng J-J, Wang DL: Chinese herbal remedy wogonin inhibits monocyte chemotactic protein-1 gene expression in human endothelial cells. *Mol Pharmacol* 2001;60:507–513.
39. Suh N, Honda T, Finlay HJ, Barchowsky A, Williams C, Benoit NE, Xie Q-W, Nathan C, Gribble GW, Sporn MB: Novel triterpenoids suppress inducible nitric oxide synthase (iNOS) and inducible cyclooxygenases (COX-2) in mouse macrophages. *Cancer Res* 1998;58:717–723.
40. Lam T, Ling T, Chowdhury C, Chao T-H, Bahjat FR, Lloyd GK, Moldawer LL, Palladino MA, Theodorakis EA: Synthesis of a novel family of diterpenes and their evaluation as anti-inflammatory agents. *Bioorg Med Chem Lett* 2003;13:3217–3221.
41. Murakami A, Takahashi D, Kinoshita T, Koshima K, Kim HW, Yoshihiro A, Nakamura Y, Jiwajinda S, Ohigashi H: Zerumbone, a Southeast Asian ginger sesquiterpene, markedly suppresses free radical generation, proinflammatory protein production, and cancer cell proliferation accompanied by apoptosis: The  $\alpha,\beta$ -unsaturated carbonyl group is a prerequisite. *Carcinogenesis* 2002;23:795–802.
42. Pelletier J-P, Yaron M, Haraoui B, Cohen P, Nahir MA, Choquette D, Wigler I, Rosner IA, Beaulieu AD: Efficacy and safety of diacerein in osteoarthritis of the knee: A double-blind, placebo-controlled trial. The Diacerein Study Group. *Arthritis Rheum* 2000;43:2339–2348.
43. Martel-Pelletier J: Pathophysiology of osteoarthritis. *Osteoarthritis Cartilage* 2003;11:1–3.
44. Frondoza CG, Frazier C, Polotsky A, Lahiji K, Hungerford DS: TNF-alpha expression of synoviocyte cultures is inhibited by hydroxy-alkoxy-phenyl compounds (HAPC) from ginger. *Trans Orthop Res Soc* 2000;46:1038.
45. Frondoza CG, Frazier C, Polotsky A, Lahiji K, Hungerford DS, Weidner M: Inhibition of chondrocyte and synoviocyte TNF- $\alpha$



- expression by hydroxy-alkoxy-phenyl compounds (HAPC) [abstract]. In: *3rd Symposium*, International Cartilage Repair Society, Gothenburg, Sweden, 2000, p. 35.
46. Frondoza CG, Sohrabi A, Polotsky A, Phan PV, Hungerford DS, Lindmark L: An in vitro screening assay for inhibitors of proinflammatory mediators in herbal extracts using human synovioocyte cultures. *In Vitro* 2004;30:95–101.
  47. Phan P, Sohrabi A, Polotsky A, Lindmark L, Hungerford DS, Lindmark L, Frondoza CG: Ginger extract components suppress induction of chemokine expression in human synovioocytes. *J Altern Complement Med* 2005;11:149–154.
  48. Stack E, DuBois RN: Regulation of cyclo-oxygenase-2. *Best Pract Res Clin Gastroenterol* 2001;15:787–800.
  49. Amin AR, Attur M, Patel RN, Thakker GD, Marshall PJ, Rediske J, Stuchin SA, Patel IR, Abramson SB: Superinduction of cyclooxygenase-2 activity in human osteoarthritis-affected cartilage. Influence of nitric oxide. *J Clin Invest* 1997;99:1231–1237.
  50. Kang RY, Freire-Moar J, Sigal E, Chu CQ: Expression of cyclooxygenase-2 in human and animal model of rheumatoid arthritis. *Br J Rheumatol* 1996;35:711–718.
  51. Surh YJ, Chun KS, Cha HH, Han SS, Keum YS, Park KK, Hahn KB: Molecular mechanisms underlying chemopreventive activities of anti-inflammatory phytochemicals: Down-regulation of COX-2 and iNOS through suppression of NF- $\kappa$ B activation. *Mutat Res* 2001;480–481:243–268.
  52. Lipsky PE: Specific COX-2 inhibitors in arthritis, oncology, and beyond: Where is the science headed? *J Rheumatol* 1999;26:25–30.
  53. Kopp E, Ghosh S: Inhibition of NF- $\kappa$ B by sodium salicylate and aspirin. *Science* 1994;265:956–959.
  54. Housby JN, Cahill CM, Chu B, Prevelige R, Bickford K, Severson MA, Calderwood SK: Non-steroidal anti-inflammatory drugs inhibit the expression of cytokines and induce HSP70 in human monocytes. *Cytokine* 1998;11:347–358.
  55. Carty TJ, Sweeney FJ, Griffiths RJ, Ernest MJ, Pillar JS, Cheng JS, Loose LD, Joseph PA, Pazoles PP, Moore PF, Nagahisa A, Kadin SB: Tenidap inhibits 5-lipoxygenase product formation in vitro, but this activity is not observed in vivo. *Inflamm Res* 1997;46:168–179.
  56. Gonzalez-Scarano F, Baltuch G: Microglia as mediators of inflammation and degenerative diseases. *Annu Rev Neurosci* 1999;22:219–240.
  57. Kreutzberg GW: Microglia: A sensor for pathological events in the CNS. *Trends Neurosci* 1996;19:312–318.
  58. Stoll G, Jander S: The role of microglia and macrophages in the pathophysiology of the CNS. *Prog Neurobiol* 1999;58:233–247.
  59. Akiyama H, Barger S, Barnum S, Bradt B, Bauer J, Cole GM, Cooper NR, Eikelboom P, Emmerling M, Fiebich BL, Finch CE, Frautschy SA, Griffin WST, Hampel H, Hull M, Landreth G, Lue L, Mrak R, Mackenzie IR, McGeer PL, O'Banion MK, Pachter J, Pasinetti G, Plata-Salaman C, Rogers J, Rydell R, Shen Y, Streit W, Strommeyer R, Tooyoma I, van Muiswinkel FL, Veerhuis R, Walker D, Webster S, Wegrzyniak B, Wenk G, Wyss-Coray T: Inflammation and Alzheimer's disease. *Neurobiol Aging* 2000;21:383–421.
  60. Orr CF, Rowe DB, Halliday GM: An inflammatory review of Parkinson's disease. *Prog Neurobiol* 2002;68:325–340.
  61. Benveniste EN: Role of macrophage/microglia in multiple sclerosis and experimental allergic encephalomyelitis. *J Mol Med* 1997;75:165–173.
  62. Lue LF, Rydell R, Brigham EF, Yang L-B, Hampel H, Murphy GM Jr, Brachova L, Yan S-D, Waker DG, Shen Y, Roger J: Inflammatory repertoire of Alzheimer's disease and nondemented elderly microglia in vitro. *Glia* 2001;35:72–79.
  63. Rozovsky I, Finch CE, Morgan TE: Age-related activation of microglia and astrocytes: In vitro studies show persistent phenotypes of aging, increased proliferation, and resistance to down-regulation. *Neurobiol Aging* 1998;19:97–103.
  64. in t' Veld BA, Ruitenbergh A, Hofman A, Launer LJ, van Duijn CM, Stijnen T, Breteler MMB, Stricker BHC: Nonsteroidal anti-inflammatory drugs and the risk of Alzheimer's disease. *N Engl J Med* 2001;345:1515–1521.
  65. Stewart WT, Kawas C, Corrada M, Metter EJ: Risk of Alzheimer's disease and duration of NSAID use. *Neurology* 1997;48:626–632.
  66. Grzanna R, Phan PV, Polotsky A, Lindmark L, Frondoza CG: Ginger extract inhibits  $\beta$ -amyloid peptide-induced cytokine and chemokine expression in cultured THP-1 monocytes. *J Altern Complement Med* 2004;10:1009–1013.
  67. Helmuth L: Protecting the brain while killing pain? *Science* 2002;297:1262–1263.
  68. Baeuerle PA, Baichwal VR: NF- $\kappa$ B as a frequent target for immunosuppressive and anti-inflammatory molecules. *Adv Immunol* 1997;65:111–137.
  69. Christman JW, Lancaster LH, Blackwell TS: Nuclear factor  $\kappa$ B: A pivotal role in the systemic inflammatory response syndrome and new target for therapy. *Intensive Care Med* 1998;24:1131–1138.
  70. Tak PP, Firestein GS: NF- $\kappa$ B: A key role in inflammatory diseases. *J Clin Invest* 2001;107:7–11.
  71. Marok R, Winyard PG, Coumbe A, Kus ML, Gaffney K, Blades S, Mapp PI, Morris CJ, Blake DR, Kaltschmidt C, Baeuerle PA: Activation transcription factor NF- $\kappa$ B in human inflamed synovial tissue. *Arthritis Rheum* 1996;39:583–591.
  72. Yamamoto Y, Gaynor RB: Therapeutic potential of inhibition of the NF- $\kappa$ B pathway in the treatment of inflammation and cancer. *J Clin Invest* 2001;107:135–142.
  73. Fakata KL: Anti-inflammatory agents for the treatment of musculoskeletal pain and arthritis. *Curr Pain Headache Rep* 2004;8:173–177.
  74. Schaible HG, Ebersberger A, von Banchet GS: Mechanisms of pain in arthritis. *Ann NY Acad Sci* 2002;966:343–354.
  75. Dedov VN, Tran VH, Duke CC, Connor M, Christie MJ, Mandadi S, Roufogalis BD: Gingerols: A novel class of vanilloid receptor (VR1) agonists. *Br J Pharmacol* 2002;137:793–798.
  76. He X, Schepelmann K, Schaible HG, Schmidt RF: Capsaicin inhibits responses of fine afferents from the knee joint of the cat to mechanical and chemical stimuli. *Brain Res* 1990;530:147–150.



**This article has been cited by:**

1. Allah Dad Talpur, Mhd Ikhwanuddin, Abol-Munafi Ambok Bolong. 2013. Nutritional effects of ginger (*Zingiber officinale* Roscoe) on immune response of Asian sea bass, *Lates calcarifer* (Bloch) and disease resistance against *Vibrio harveyi*. *Aquaculture* . [CrossRef]
2. R. Jaffe Cardioprotective Nutrients 103-119. [CrossRef]
3. A.N. Prabhu, A.R. Shivashankara, R. Haniadka, P.L. Palatty, D. Prabhu, M.S. Baliga Antiatherogenic Effects of Ginger (*Zingiber officinale* Roscoe) 693-704. [CrossRef]
4. M.S. Baliga, L. Latheef, R. Haniadka, F. Fazal, J. Chacko, R. Arora Ginger (*Zingiber officinale* Roscoe) in the Treatment and Prevention of Arthritis 529-544. [CrossRef]
5. How to use the monographs 353-961. [CrossRef]
6. Kainat Khan, Akanksha Singh, Monika Mittal, Kunal Sharan, Nidhi Singh, Preety Dixit, Sabyasachi Sanyal, Rakesh Maurya, Naibedya Chattopadhyay. 2012. [6]-Gingerol induces bone loss in ovary intact adult mice and augments osteoclast function via the transient receptor potential vanilloid 1 channel. *Molecular Nutrition & Food Research* 56:12, 1860-1873. [CrossRef]
7. Marie-Lou Gauthier, Francis Beaudry, Pascal Vachon. 2012. Intrathecal [6]-Gingerol Administration Alleviates Peripherally induced Neuropathic Pain in Male Sprague-Dawley Rats. *Phytotherapy Research* n/a-n/a. [CrossRef]
8. I. Rahath Kubra, L. Jagan Mohan Rao. 2012. An Impression on Current Developments in the Technology, Chemistry, and Biological Activities of Ginger (*Zingiber officinale* Roscoe). *Critical Reviews in Food Science and Nutrition* 52:8, 651-688. [CrossRef]
9. Sang Keun Ha, Eunjung Moon, Mi Sun Ju, Dong Hyun Kim, Jong Hoon Ryu, Myung Sook Oh, Sun Yeou Kim. 2012. 6-Shogaol, a ginger product, modulates neuroinflammation: A new approach to neuroprotection. *Neuropharmacology* 63:2, 211-223. [CrossRef]
10. Vladimir N. Drozdov, Victoria A. Kim, Elena V. Tkachenko, Galina G. Varvanina. 2012. Influence of a Specific Ginger Combination on Gastropathy Conditions in Patients with Osteoarthritis of the Knee or Hip. *The Journal of Alternative and Complementary Medicine* 18:6, 583-588. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
11. Sehwan Shim, Jungkee Kwon. 2012. Effects of [6]-shogaol on cholinergic signaling in HT22 cells following neuronal damage induced by hydrogen peroxide. *Food and Chemical Toxicology* 50:5, 1454-1459. [CrossRef]
12. Ghanshyam B. Dudhatra, Shailesh K. Mody, Madhavi M. Awale, Hitesh B. Patel, Chirag M. Modi, Avinash Kumar, Divyesh R. Kamani, Bhavesh N. Chauhan. 2012. A Comprehensive Review on Pharmacotherapeutics of Herbal Bioenhancers. *The Scientific World Journal* 2012, 1-33. [CrossRef]
13. Tullayakorn Plengsuriyakarn, Vithoon Viyanant, Veerachai Eursitthichai, Pornthipa Picha, Piengchai Kupradinant, Arunporn Itharat, Kesara Na-Bangchang. 2012. Anticancer activities against cholangiocarcinoma, toxicity and pharmacological activities of Thai medicinal plants in animal models. *BMC Complementary and Alternative Medicine* 12:1, 23. [CrossRef]
14. Alois Jungbauer, Svjetlana Medjakovic. 2012. Anti-inflammatory properties of culinary herbs and spices that ameliorate the effects of metabolic syndrome. *Maturitas* . [CrossRef]
15. Manjeshwar Shrinath Baliga, Raghavendra Haniadka, Manisha Maria Pereira, Karadka Ramdas Thilakchand, Suresh Rao, Rajesh Arora. 2012. Radioprotective effects of *Zingiber officinale* Roscoe (Ginger): past, present and future. *Food & Function* . [CrossRef]
16. Søren Ribel-Madsen, Else Marie Bartels, Anders Stockmarr, Arne Borgwardt, Claus Cornett, Bente Danneskiold-Samsøe, Henning Bliddal. 2012. A Synoviocyte Model for Osteoarthritis and Rheumatoid Arthritis: Response to Ibuprofen, Betamethasone, and Ginger Extract—A Cross-Sectional In Vitro Study. *Arthritis* 2012, 1-9. [CrossRef]
17. Yueh-Ping Chang, Chun-Hung Liu, Chih-Chung Wu, Chi-Ming Chiang, Juang-Lin Lian, Shu-Ling Hsieh. 2011. Dietary administration of zingerone to enhance growth, non-specific immune response, and resistance to *Vibrio alginolyticus* in Pacific white shrimp (*Litopenaeus vannamei*) juveniles. *Fish & Shellfish Immunology* . [CrossRef]
18. Mohsen Zahmatkash, Mohammad Reza Vafaenasa. 2011. Comparing Analgesic Effects of a Topical Herbal Mixed Medicine with Salicylate in Patients with Knee Osteoarthritis. *Pakistan Journal of Biological Sciences* 14:13, 715-719. [CrossRef]
19. Y. Amarnath Reddy, M. Chalamaiah, B. Ramesh, G. Balaji, P. Indira. 2011. Ameliorating activity of ginger (*Zingiber officinale*) extract against lead induced renal toxicity in male rats. *Journal of Food Science and Technology* . [CrossRef]
20. Xiao-Hong Li, Kristine C-Y McGrath, Srinivas Nammi, Alison K. Heather, Basil D. Roufogalis. 2011. Attenuation of Liver Pro-Inflammatory Responses by *Zingiber officinale* via Inhibition of NF-kappa B Activation in High-Fat Diet-Fed Rats. *Basic & Clinical Pharmacology & Toxicology* no-no. [CrossRef]

21. Gamal Ramadan, Mohammed Ali Al-Kahtani, Wael Mohamed El-Sayed. 2011. Anti-inflammatory and Anti-oxidant Properties of *Curcuma longa* (Turmeric) Versus *Zingiber officinale* (Ginger) Rhizomes in Rat Adjuvant-Induced Arthritis. *Inflammation* 34:4, 291-301. [[CrossRef](#)]
22. Sehwan Shim, Sokho Kim, Dea-Seung Choi, Young-Bae Kwon, Jungkee Kwon. 2011. Anti-inflammatory effects of [6]-shogaol: Potential roles of HDAC inhibition and HSP70 induction. *Food and Chemical Toxicology* . [[CrossRef](#)]
23. Anjali B. Ganjare, Sunil A. Nirmal, Ruksana A. Rub, Anuja N. Patil, Shashikant R. Pattan. 2011. Use of *Cordia dichotoma* bark in the treatment of ulcerative colitis. *Pharmaceutical Biology* 49:8, 850-855. [[CrossRef](#)]
24. Bharati B. Zaware, Sunil A. Nirmal, D. G. Baheti, Anuja N. Patil, Subhash C. Mandal. 2011. Potential of *Vitex negundo* roots in the treatment of ulcerative colitis in mice. *Pharmaceutical Biology* 49:8, 874-878. [[CrossRef](#)]
25. Manjeshwar Shrinath Baliga, Raghavendra Haniadka, Manisha Maria Pereira, Jason Jerome D'Souza, Princy Louis Pallaty, Harshith P. Bhat, Sandhya Popuri. 2011. Update on the Chemopreventive Effects of Ginger and its Phytochemicals. *Critical Reviews in Food Science and Nutrition* 51:6, 499-523. [[CrossRef](#)]
26. Anjali B. Ganjare, Sunil A. Nirmal, Anuja N. Patil. 2011. Use of apigenin from *Cordia dichotoma* in the treatment of colitis. *Fitoterapia* . [[CrossRef](#)]
27. Jung-Min Yon, In-Jeoung Baek, Beom Jun Lee, Young Won Yun, Sang-Yoon Nam. 2011. Emodin and [6]-gingerol lessen hypoxia-induced embryotoxicities in cultured mouse whole embryos via upregulation of hypoxia-inducible factor 1 $\alpha$  and intracellular superoxide dismutases. *Reproductive Toxicology* 31:4, 513-518. [[CrossRef](#)]
28. Soon-Yong Choi, Gil-Soon Park, Sung Yoon Lee, Ji Yeon Kim, Young Kook Kim. 2011. The conformation and CETP inhibitory activity of [10]-dehydrogingerdione isolated from *Zingiber officinale*. *Archives of Pharmacal Research* 34:5, 727-731. [[CrossRef](#)]
29. Xiao Wang, Zhenjia Zheng, Xingfeng Guo, Jinpeng Yuan, Chengchao Zheng. 2011. Preparative separation of gingerols from *Zingiber officinale* by high-speed counter-current chromatography using stepwise elution. *Food Chemistry* 125:4, 1476-1480. [[CrossRef](#)]
30. Masood Sadiq Butt, M. Tauseef Sultan. 2011. Ginger and its Health Claims: Molecular Aspects. *Critical Reviews in Food Science and Nutrition* 51:5, 383-393. [[CrossRef](#)]
31. Ann Bode, Zigang Dong The Amazing and Mighty Ginger 2011 5386, 131-156. [[CrossRef](#)]
32. Peraphan Pothacharoen, Kanyamas Choocheep, Thanyaluck Phitak, Wilart Pompimon, Prachya Kongtawelert. 2011. *Alpinia galanga* extracts downregulate interleukin-1 $\beta$ -induced matrix metalloproteinases expression in human synovial fibroblasts. *In Vitro Cellular & Developmental Biology - Animal* 47:3, 183-187. [[CrossRef](#)]
33. D.N.A Tagoe, H.D. Nyarko, R. Akpaka. 2011. A Comparison of the Antifungal Properties of Onion (*Allium cepa*), Ginger (*Zingiber officinale*) and Garlic (*Allium sativum*) against *Aspergillus flavus*, *Aspergillus niger* and *Cladosporium herbarum*. *Research Journal of Medicinal Plant* 5:3, 281-287. [[CrossRef](#)]
34. Flora F. Barsoum. 2011. Synthesis and Molecular Modeling Studies of Anti-inflammatory Active 1H-Pyrrolizine-5-carboxamides. *Archiv der Pharmazie* 344:1, 56-65. [[CrossRef](#)]
35. Yasuko Sone, Joon-Kwan Moon, Truong Tuyet Mai, Nghiem Nguyet Thu, Eri Asano, Keiko Yamaguchi, Yuzuru Otsuka, Takayuki Shibamoto. 2011. Antioxidant/anti-inflammatory activities and total phenolic content of extracts obtained from plants grown in Vietnam. *Journal of the Science of Food and Agriculture* n/a-n/a. [[CrossRef](#)]
36. Monica Borgatti, Irene Mancini, Nicoletta Bianchi, Alessandra Guerrini, Ilaria Lampronti, Damiano Rossi, Gianni Sacchetti, Roberto Gambari. 2011. Bergamot (*Citrus bergamia* Risso) fruit extracts and identified components alter expression of interleukin 8 gene in cystic fibrosis bronchial epithelial cell lines. *BMC Biochemistry* 12:1, 15. [[CrossRef](#)]
37. Kelly Galvin, Madelaine Bishop Integumentary system 565-623. [[CrossRef](#)]
38. Kelly Galvin, Madelaine Bishop Gastrointestinal system 43-128. [[CrossRef](#)]
39. Jeffrey K. Actor Herbal Medicines with Immunomodulatory Effects 73-124. [[CrossRef](#)]
40. Keith Singletary. 2010. Ginger. *Nutrition Today* 45:4, 171-183. [[CrossRef](#)]
41. Kim Edward LeBlanc, Leanne L. LeBlanc. 2010. Musculoskeletal Disorders. *Primary Care: Clinics in Office Practice* 37:2, 389-406. [[CrossRef](#)]
42. Ademola A. Oyagbemi, Adebowale B. Saba, Odunayo I. Azeez. 2010. Molecular targets of [6]-gingerol: Its potential roles in cancer chemoprevention. *BioFactors* 36:3, 169-178. [[CrossRef](#)]
43. Evan Prince Sabina, MahaboobKhan Rasool, Lazar Mathew, Panneerselvam EzilRani, Haridas Indu. 2010. 6-Shogaol inhibits monosodium urate crystal-induced inflammation – An in vivo and in vitro study. *Food and Chemical Toxicology* 48:1, 229-235. [[CrossRef](#)]

44. Simla Basar, Klaus Uhlenhut, Petra Häfner, Florian Schöne, Johannes Westendorf. 2010. Analgesic and antiinflammatory activity of *Morinda citrifolia* L. (Noni) fruit. *Phytotherapy Research* **24**:1, 38-42. [[CrossRef](#)]
45. Hiroshi UEDA, Katsunari IPPOUSHI, Atsuko TAKEUCHI. 2010. Repeated Oral Administration of a Squeezed Ginger (*Zingiber officinale*) Extract Augmented the Serum Corticosterone Level and Had Anti-Inflammatory Properties. *Bioscience, Biotechnology, and Biochemistry* **74**:11, 2248-2252. [[CrossRef](#)]
46. Sudesh Agrawal, Amitabha Chakrabarti Potential Nutraceutical Ingredients from Plant Origin 27-68. [[CrossRef](#)]
47. E J Nya, B Austin. 2009. Use of dietary ginger, *Zingiber officinale* Roscoe, as an immunostimulant to control *Aeromonas hydrophila* infections in rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of Fish Diseases* **32**:11, 971-977. [[CrossRef](#)]
48. Chia-Feng Kuo, Ming-Hon Hou, Tsu-Shing Wang, Charng-Cherng Chyau, Yi-Ting Chen. 2009. Enhanced antioxidant activity of *Monascus pilosus* fermented products by addition of ginger to the medium. *Food Chemistry* **116**:4, 915-922. [[CrossRef](#)]
49. Bill Roschek, Ryan C. Fink, Matthew McMichael, Randall S. Alberte. 2009. Nettle extract (*Urtica dioica*) affects key receptors and enzymes associated with allergic rhinitis. *Phytotherapy Research* **23**:7, 920-926. [[CrossRef](#)]
50. Hyo Won Jung, Cheol-Ho Yoon, Kwon Moo Park, Hyung Soo Han, Yong-Ki Park. 2009. Hexane fraction of *Zingiberis Rhizoma Crudus* extract inhibits the production of nitric oxide and proinflammatory cytokines in LPS-stimulated BV2 microglial cells via the NF-kappaB pathway. *Food and Chemical Toxicology* **47**:6, 1190-1197. [[CrossRef](#)]
51. Giti Ozgoli, Marjan Goli, Fariborz Moattar. 2009. Comparison of Effects of Ginger, Mefenamic Acid, and Ibuprofen on Pain in Women with Primary Dysmenorrhea. *The Journal of Alternative and Complementary Medicine* **15**:2, 129-132. [[Abstract](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
52. Chris D. Meletis, Nieske Zabriskie, Robert Rountree. 2009. Identifying and Treating Lyme Disease. *Alternative and Complementary Therapies* **15**:1, 17-23. [[Citation](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
53. Apollinaire Tsopmo, Pierre Kamnaing, Jean Watchueng, Jin-Ming Gao, Yasuo Konishi, Olov Sterner. 2009. Chemical constituents from the bark of *Anisopus mannii*. *Canadian Journal of Chemistry* **87**:2, 397-400. [[CrossRef](#)]
54. Herbal Medicines – Acanthaceae – Zygophyllaceae 31-235. [[CrossRef](#)]
55. Abdel-Motaal M. Fouda, Mohamed Y. Berika. 2009. Evaluation of the Effect of Hydroalcoholic Extract of *Zingiber officinale* Rhizomes in Rat Collagen-induced Arthritis. *Basic & Clinical Pharmacology & Toxicology* **104**:3, 262. [[CrossRef](#)]
56. Thanyaluck Phitak, Kanyamas Choocheep, Peraphan Pothacharoen, Wilart Pompimon, Bhusana Premanode, Prachya Kongtawelert. 2009. The effects of p-hydroxycinnamaldehyde from *Alpinia galanga* extracts on human chondrocytes. *Phytochemistry* **70**:2, 237-243. [[CrossRef](#)]
57. Marie Louise Berthe Ahui, Pierre Champy, Abdulraouf Ramadan, Linh Pham Van, Luiza Araujo, Konan Brou André, Séverine Diem, Diane Damotte, Séraphin Kati-Coulibaly, Michel Atté Offoumou. 2008. Ginger prevents Th2-mediated immune responses in a mouse model of airway inflammation. *International Immunopharmacology* **8**:12, 1626-1632. [[CrossRef](#)]
58. Suzanna M. Zick, Mack T. Ruffin, Julia Lee, Daniel P. Normolle, Rivka Siden, Sara Alrawi, Dean E. Brenner. 2008. Phase II trial of encapsulated ginger as a treatment for chemotherapy-induced nausea and vomiting. *Supportive Care in Cancer* . [[CrossRef](#)]
59. C WILLIAMS, E LAMPRECHT. 2008. Some commonly fed herbs and other functional foods in equine nutrition: A review. *The Veterinary Journal* **178**:1, 21-31. [[CrossRef](#)]
60. H ELABHAR, L HAMMAD, H GAWAD. 2008. Modulating effect of ginger extract on rats with ulcerative colitis. *Journal of Ethnopharmacology* **118**:3, 367-372. [[CrossRef](#)]
61. B ALI, G BLUNDEN, M TANIRA, A NEMMAR. 2008. Some phytochemical, pharmacological and toxicological properties of ginger (*Zingiber officinale* Roscoe): A review of recent research. *Food and Chemical Toxicology* **46**:2, 409-420. [[CrossRef](#)]
62. Rheumatoid Arthritis 339-352. [[Citation](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
63. Chris D. Meletis, Nieske L. Zabriskie, Robert Rountree Clinical Natural Medicine Handbook . [[Abstract](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)] [[Supplemental Material](#)]
64. Natural Approaches to Controlling Inflammatory Disease 229-239. [[Citation](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
65. M.H. Pittler, E. Ernst 48 Treatments used in complementary and alternative medicine **30**, 551-560. [[CrossRef](#)]
66. Mari Maeda-Yamamoto, Kaori Ema, Ikuo Shibuichi. 2007. In vitro and in vivo anti-allergic effects of 'benifuuki' green tea containing O-methylated catechin and ginger extract enhancement. *Cytotechnology* **55**:2-3, 135-142. [[CrossRef](#)]
67. Amanat Ali, Anwarul Hassan Gilani. 2007. Medicinal Value of Ginger with Focus on its Use in Nausea and Vomiting of Pregnancy. *International Journal of Food Properties* **10**:2, 269-278. [[CrossRef](#)]
68. S TRIPATHI, K MAIER, D BRUCH, D KITTUR. 2007. Effect of 6-Gingerol on Pro-Inflammatory Cytokine Production and Costimulatory Molecule Expression in Murine Peritoneal Macrophages. *Journal of Surgical Research* **138**:2, 209-213. [[CrossRef](#)]



69. Peraphan Pothacharoen, Kanyamas Choocheep, Tanyaluck Pitak, Wilart Pompimon, Bhusana Premanode, Timothy E. Hardingham, Prachya Kongtawelert. 2006. Effect of *Alpinia galanga* extract on cartilage degradation and on gene expression in human chondrocyte and synovial fibroblast metabolism. *Central European Journal of Biology* 1:3, 430-450. [[CrossRef](#)]
70. Mou-Tuan Huang, Yue Liu, Divya Ramji, Chih-Yu Lo, Geetha Ghai, Slavik Dushenkov, Chi-Tang Ho. 2006. Inhibitory effects of black tea theaflavin derivatives on 12-O-tetradecanoylphorbol-13-acetate-induced inflammation and arachidonic acid metabolism in mouse ears. *Molecular Nutrition & Food Research* 50:2, 115-122. [[CrossRef](#)]
71. Masako ABE, Yoshio OZAWA, Yasushi UDA, Yasujiro MORIMITSU, Yoshimasa NAKAMURA, Toshihiko OSAWA. 2006. A Novel Labdane-Type Trialdehyde from Myoga (*Zingiber mioga* Roscoe) That Potently Inhibits Human Platelet Aggregation and Human 5-Lipoxygenase. *Bioscience, Biotechnology, and Biochemistry* 70:10, 2494-2500. [[CrossRef](#)]
72. 2005. Literature Watch. *Alternative and Complementary Therapies* 11:5, 273-274. [[Citation](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
73. Mari Maeda-Yamamoto, Hiroshi Nagai, Kaori Ema, Emi Kanda, Norihisa Okada, Masaaki Yasue. 2005. Effects and Safety of Consecutive Intake of Benifuuki Green Tea and Enhancement of the Effect by Ginger Extract in Subjects with Japanese Cedar-pollinosis. *Nippon Shokuhin Kagaku Kogaku Kaishi* 52:12, 584-593. [[CrossRef](#)]